

CHROM. 14,088

Note

Determination of 7-dehydrocholesterol in rat skin and liver by high-performance liquid chromatography

K. TAKADA*, R. ITO, K. TOKUNAGA, T. KOBAYASHI and T. TAKAO

Department of Hygienic Chemistry, Kobe Women's College of Pharmacy, Motoyamakita-machi, Higashi-nada-ku, Kobe 658 (Japan)

(Received May 12th, 1981)

7-Dehydrocholesterol (7-DHC) is not only an intermediate of cholesterol biogenesis but is also a photosynthetic precursor of vitamin D₃, *i.e.*, provitamin D₃ (ref. 1).

Conventional methods used for the determination of 7-DHC in animal tissues are spectrophotometric methods, including fractionation of unsaponifiable matter by means of column chromatography² or thin-layer chromatography³. However, the methods so far reported were neither accurate nor applicable to liver, owing to irrelevant UV-absorbing impurities and low sensitivity. In order to investigate the metabolism of provitamin D₃ in detail, a simple, precise and sensitive method for the determination of 7-DHC in body tissues was required. This paper reports a method for the determination of 7-DHC in rat skin and liver by high-performance liquid chromatography (HPLC).

EXPERIMENTAL

Chemicals

Crystalline 7-DHC was purchased from Fluka (Buchs, Switzerland) and was recrystallized from diethyl ether (m.p. 150–151°C). Its purity was checked by HPLC.

All solvents were purified according to conventional methods in our laboratory and were redistilled before use. Other reagents were of analytical-reagent grade.

Apparatus

Two HPLC systems were used. The system used for the pre-purification of unsaponifiable matter obtained from rat liver consisted of a Shimadzu–DuPont LC 841 high-performance liquid chromatograph, equipped with a UV detector (254 nm) (Shimadzu, Kyoto, Japan), a 25 cm × 6.2 mm I.D. Zorbax SIL column (DuPont, Wilmington, DE, U.S.A.) and a Model SIL-1A LC injector (Shimadzu). The other system, used for the determination of 7-DHC in the unsaponifiable matter of rat skin or in the pre-purified 7-DHC fraction of rat liver, consisted of a Shimadzu LC 3A high-performance liquid chromatograph, a 25 cm × 4.6 mm I.D. Zorbax SIL column, and a Model UVD-2 UV detector (254 nm; maximum sensitivity $1 \cdot 10^{-3}$ absorbance unit) (Shimadzu).

A Gilson Micro Fractionator (Gilson Instrument Co., Middleton, WI, U.S.A.) was connected to the first chromatography system to fractionate the eluate.

UV absorption spectra were recorded with a Hitachi 323 automatic spectrophotometer (Hitachi, Tokyo, Japan).

Assay of 7-DHC in skin

A piece of dorsal rat skin (1 g) was cut into small pieces and saponified by refluxing for 30 min with a mixture of 25 ml of ethanol, 10 ml of 20% pyrogallol solution in ethanol and 4 ml of 85% (w/v) potassium hydroxide solution. The unsaponifiable matter was extracted with 50 ml of benzene according to the method of Mulder and Vries⁴. After the separated benzene layer had been filtered through a Whatman IPS phase-separating paper, 2 ml of the filtrate were dried under a stream of nitrogen and the residue was dissolved in 500 μ l of *n*-hexane. A 50- μ l volume of the solution was applied to a Zorbax SIL column (25 cm \times 4.6 mm I.D.), and was eluted at a flow-rate of 1.8 ml/min with 0.5% 2-propanol in *n*-hexane. The sensitivity of the UV detector was set at $32 \cdot 10^{-3}$ absorbance units full-scale (a.u.f.s.).

Assay of 7-DHC in liver

Rat liver (5–10 g) was cut into small pieces, saponified and extracted as above, but with double the amounts of reagents. The residue was dissolved in 100 μ l of *n*-hexane and 50 μ l of the solution were subjected to HPLC for the pre-purification of the unsaponifiable matter, using a Zorbax SIL column (25 cm \times 6.2 mm I.D.) with a flow-rate of 1.6 ml/min of 1.0% 2-propanol in *n*-hexane at a pressure of 60–80 kg/cm². The eluate corresponding to the 7-DHC fraction (20–24 min) was collected with the fraction collector and was subsequently applied to analytical HPLC on a Zorbax SIL column (25 cm \times 4.6 mm I.D.) under the same conditions as for the assay of 7-DHC in skin.

As the content of 7-DHC in the liver is much smaller than that in the skin, the sensitivity of the UV detector was set to the maximum ($1 \cdot 10^{-3}$ a.u.f.s.).

RESULTS AND DISCUSSION

A piece of dorsal skin obtained from an adult male Wistar rat was treated according to the method described above and a typical chromatogram of the unsaponifiable matter is illustrated in Fig. 1. The 7-DHC peak was clearly separated from peaks of unknown material. The chromatogram shows that 7-DHC is a major UV-absorbing substance in the unsaponifiable matter of rat skin.

Peak identification was carried out on the eluate fraction corresponding to the peak of 7-DHC by means of reversed-phase HPLC on a Nucleosil 5C₁₈ column (30 cm \times 7.5 mm I.D.) eluted with methanol–2-propanol–water (4:2:1) at a flow-rate of 1.7 ml/min and UV spectrophotometry.

The overall recovery of 100 μ g of 7-DHC added to the sample was $107.3 \pm 3.4\%$ [mean \pm standard deviation (S.D.), $n = 6$].

Application of the same procedure to the determination of 7-DHC in liver was not successful, owing to extensive peak overlap with impurities. The content of 7-DHC in rat liver was too small to quantify directly from the crude unsaponifiable matter. In order to eliminate the UV-absorbing impurities, the unsaponifiable matter

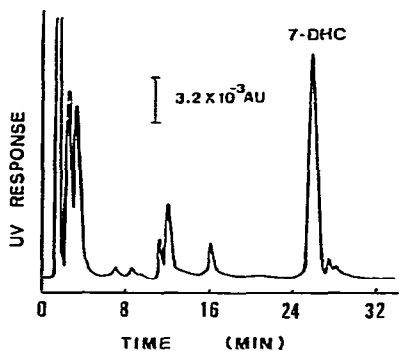


Fig. 1. Analysis of unsaponifiable matter obtained from rat skin by HPLC on a Zorbax SIL column (25 cm \times 4.6 mm I.D.) with a flow-rate of 1.8 ml/min of 0.5% 2-propanol in *n*-hexane.

was first subjected to semi-preparative HPLC on a Zorbax SIL column (25 cm \times 6.2 mm I.D.). Fig. 2a shows a typical chromatogram of the unsaponifiable matter obtained from rat liver. The eluate corresponding to the 7-DHC fraction (represented by arrows) was subsequently subjected to analytical HPLC under the same conditions as for the assay of 7-DHC in skin. Although baseline separation from unknown peaks could not be obtained, an adequate and reproducible separation was achieved (Fig. 2b). The overall recovery of 1 μ g of 7-DHC added to the sample was $95.0 \pm 5.6\%$ (mean \pm S.D., $n = 6$).

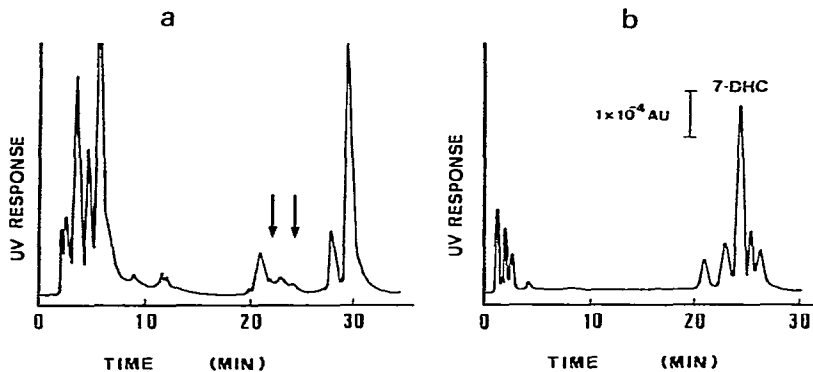


Fig. 2. (a) Semi-preparative HPLC of unsaponifiable matter obtained from rat liver on a Zorbax SIL column (25 cm \times 6.2 mm I.D.) with a flow-rate of 1.6 ml/min of 1.0% 2-propanol in *n*-hexane. Arrows indicate the beginning and end of elution of the 7-DHC fraction. (b) Analytical HPLC of the pre-purified 7-DHC fraction obtained by semi-preparative HPLC. The conditions were identical with those for the analysis of skin unsaponifiable matter.

The peak corresponding to 7-DHC was identified as follows. A large amount of pre-purified 7-DHC fraction was collected from the other parts of the unsaponifiable matter and was subjected once more to semi-preparative HPLC. The 7-DHC fraction obtained was further purified by subjecting it to analytical HPLC. The eluate corresponding to the 7-DHC peak was collected and then the UV absorption spectrum was recorded. As Fig. 3 shows, the spectrum gave the characteristic four absor-

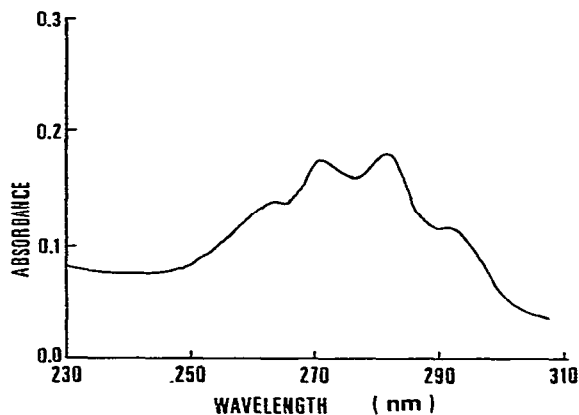


Fig. 3. Ultraviolet spectrum of purified 7-DHC fraction obtained from rat liver.

TABLE I
COMPARISON OF 7-DHC CONTENTS IN RAT SKIN AND LIVER

Sample No.	7-DHC content ($\mu\text{g/g}$)	
	Skin	Liver
1	274	1.88
2	207	1.10
3	256	0.64
4	262	0.74
5	280	1.25
6	235	0.96
7	210	1.17
8	256	0.79
Mean \pm S.D.	248 \pm 26	1.07 \pm 0.37

bance maxima for a 5,7-diene sterol such as 7-DHC. The isolation and identification of 7-DHC from rat liver have not been accomplished before by HPLC.

These methods were then applied to a comparative study of the levels of 7-DHC in rat skin and liver. Table I shows the 7-DHC levels found in the skins and livers of adult male Wistar rats (8 weeks old). As suggested before², the 7-DHC contents of the liver were very small in comparison with the skin contents (*ca.* 1:250). The high level of 7-DHC in the skin would be adequate for the photosynthesis of vitamin D₃.

REFERENCES

- 1 H. F. DeLuca, J. W. Blunt and H. Rikkers, *The Vitamins*, Vol. 3, Academic Press, New York, 2nd ed., 1971, pp. 213-230.
- 2 M. Glover, J. Glover and R. A. Morton, *Biochemistry*, 51 (1952) 1.
- 3 B. Doboszynska and E. Burakowska, *Bull. Acad. Pol. Sci., Ser. Sci. Biol.*, 19 (1971) 553.
- 4 F. J. Mulder and H. Vries, *J. Ass. Offic. Anal. Chem.*, 52 (1969) 1189.